



中国认可
国际互认
检测
TESTING
CNAS L1225



TEST REPORT

Report No.: 4478420080329R1

Product Name: surgical mask

Product Model: non-sterile

BRAND: _____

Applicant: New Island Printing (Liaoning) Co., Ltd




CHINA CERTIFICATION & INSPECTION GROUP SHENZHEN CO., LTD.
SHENZHEN HUATONGWEI INTERNATIONAL INSPECTION CO.,LTD.





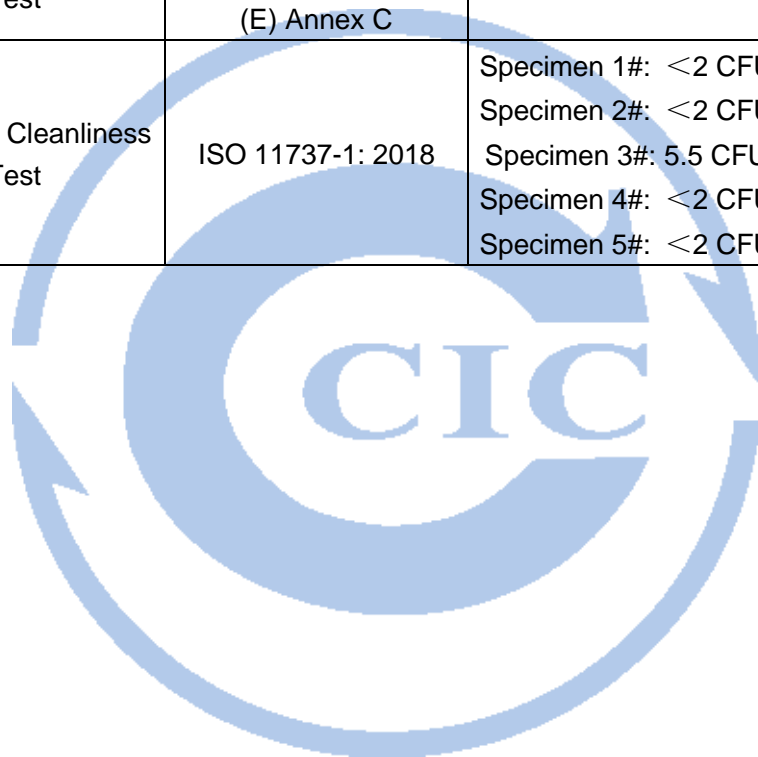
Test Report

APPLICANT	New Island Printing (Liaoning) Co., Ltd		
ADDRESS	No.17 Xinmin Street, West District, Yingkou City, Liaoning Province, China		
SAMPLE NAME	surgical mask		
SAMPLE MODEL	non-sterile		
SAMPLE QUANTITY	80pcs		
TYPE	Type I		
TEST REQUESTED	As specified by client, for details refer to next page(s).		
TEST METHOD	Please refer to next page.		
TEST RESULTS	Please refer to next page(s).		
RECEIVED DATE	08 17, 2020	TESTING PERIOD	08 17, 2020~09 08, 2020
SAMPLE PHOTO:	1-0 		



Summary of Test Results

No.	Test Item	Test Standard	Test Result	Judgement
1	Bacterial Filtration Efficiency (BFE) Test	EN 14683:2019+AC:2019 (E) Annex B	Specimen 1#: 99.0% Specimen 2#: 98.5% Specimen 3#: 98.7% Specimen 4#: 98.2% Specimen 5#: 98.0%	Pass
2	Differential Pressure Test	EN 14683:2019+AC:2019 (E) Annex C	Max.:39.4Pa/cm ²	Pass
3	*Microbial Cleanliness Test	ISO 11737-1: 2018	Specimen 1#: <2 CFU/g Specimen 2#: <2 CFU/g Specimen 3#: 5.5 CFU/g Specimen 4#: <2 CFU/g Specimen 5#: <2 CFU/g	Pass





Bacterial Filtration Efficiency (BFE) Test

1. Purpose

For evaluating the bacterial filtration efficiency (BFE) of mask.

2. Sample description was given by client

Sample description: Single-use surgical mask with ear loop

Lot Number : /

Sample Receiving Date :2020-08-17

3. Test Method

EN 14683:2019+AC:2019(E) Annex B

4. Apparatus and materials

- 4.1 *Staphylococcus aureus* ATCC 6538.
- 4.2 Peptone water.
- 4.3 Tryptic Soy Broth(TSB).
- 4.4 Tryptic Soy Agar(TSA).
- 4.5 Bacterial filtration efficiency test apparatus.
- 4.6 Six-stage viable particle Anderson sampler.
- 4.7 Flow meters.

5. Test specimen

5.1 As requested by client, take a total of 5 test specimens.

Prior to testing, condition all test specimens for a minimum of 4 h at $(21\pm 5)^{\circ}\text{C}$ and $(85\pm 5)\%$ relative humidity.



1. Procedure

- 1.1 Preparation of the bacterial challenge: Dilute the culture in peptone water to achieve a concentration of approximately 5×10^5 CFU/mL.
- 1.2 Adjust the flow rate through the Anderson sampler to 28.3 L/min.
- 1.3 Deliver the challenge to the nebulizer using a syringe pump. Purge tubing and nebulizer of air bubbles.
- 1.4 Perform a positive control run without a test specimen to determine the number of viable aerosol particles being generated. The mean particle size (MPS) of the aerosol will also be calculated from the results of these positive control plates.
 - 1.4.1 Initiate the aerosol challenge by turning on the air pressure and pump connected to the nebulizer. Immediately begin sampling the aerosol using the Anderson sampler.
 - 1.4.2 Time the challenge suspension to be delivered to the nebulizer for 1 min.
 - 1.4.3 Time the air pressure and Anderson sampler to run for 2 min.
 - 1.4.4 At the conclusion of the positive control run, remove plates from the Anderson sampler.
- 1.5 Place new agar plates into Anderson sampler and clamp the test specimen into the top of the Anderson sampler, with the inside of the specimen facing towards the bacterial challenge (test area: 77cm^2).
- 1.6 Repeat the challenge procedure for each test specimen.
- 1.7 Repeat a positive control after completion of the sample set.
- 1.8 Perform a negative control run by collecting a 2 min sample of air from the aerosol chamber. No bacterial challenge should be pumped into the nebulizer during the collection of the negative control.
- 1.9 Incubate agar plates at $(37 \pm 2)^\circ\text{C}$ for (20 to 52) h.
- 1.10 Count each of the six-stage plates of the Anderson sampler.

2. Calculation

Total the count from each of the six plates for the test specimens and positive controls, as specified by the manufacture of Anderson sampler. The filtration efficiency percentages are calculated as follows:

$$\text{BFE} = (C - T) / C \times 100$$

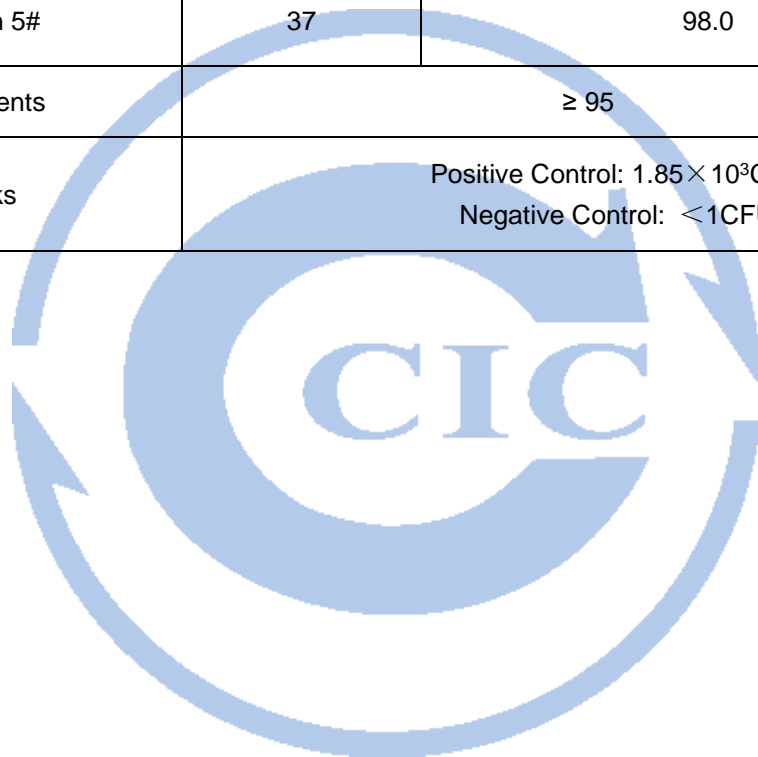
T is the total plate count for the test specimen.

C is the mean of the total plate counts for the two positive controls.



8.Test results

Specimens	T	Results (%)
Specimen 1#	18	99.0
Specimen 2#	28	98.5
Specimen 3#	24	98.7
Specimen 4#	33	98.2
Specimen 5#	37	98.0
Requirements		≥ 95
Remarks		Positive Control: 1.85×10^3 CFU Negative Control: <1CFU



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Differential pressure Test

1. Purpose

The purpose of the test was to measure the differential pressure of masks.

2. Sample description was given by client

Sample description: Single-use surgical mask with ear loop

Lot Number : /

Sample Receiving Date :2020-08-17

3. Test Method

EN 14683:2019+AC:2019(E) Annex C

4. Apparatus and materials

Differential pressure testing instrument

5. Test specimen

5.1 Test specimen are complete masks or shall be cut from masks. Each specimen shall be able to provide 5 different circular test areas of 2.5 cm in diameter.

5.2 Prior to testing, condition all test specimens for a minimum of 4 h at (21±5) °C and (85±5)% relative humidity.

6. Procedure

6.1 Without a specimen in place, the holder is closed and the differential manometer is zeroed.

The pump is started and the flow of air adjusted to 8 L/min.

6.2 The pretreated specimen is placed across the orifice (total area 4.9cm², test area diameter 25mm) and clamped into place so as to minimize air leaks.

6.3 Due to the presence of an alignment system the tested area of the specimen should be perfectly in line and across the flow of air.

6.4 The differential pressure is read directly.

6.5 The procedure described in steps 6.1-6.4 is carried out on 5 different areas of the mask and readings averaged.

Results:

Specimen	Test Results (Pa/cm ²)	Maximum (Pa/cm ²)	Requirements	Judgement
1#	38.4	39.4	< 40	Pass
2#	39.4			
3#	39.2			
4#	38.2			
5#	38.0			



Microbial Cleanliness Test

1.0 Purpose

To describe the population of viable microorganisms present on or in product

2.0 Reference

Medical face masks - Requirements and test methods EN 14683-2019

Sterilization of medical devices – Microbiological methods – Part 1: Determination of a population of microorganisms on products ISO 11737-1: 2018

3.0 Equipment and reagents

3.1 Instruments

Vertical pressure steam sterilizer (SHB026), Steel Straight Scale (SHB076), Electronic Balance (SHB016), Clean bench (SHB015), Bench type low speed large capacity centrifuge (SHB021), Inverted microscope (SHB005), Constant Temperature Vibrator (SHB007), Biochemical incubator(SHB024), Biochemical incubator(SHB025)

3.2 Reagents

SDA (Beijing Land Bridge Technology Co., Ltd, Lot No:20190912) ,TSA (HOPEBIO, Lot No:20190 613) ,Sodium chloride-peptone buffer (Beijing Land Bridge Technology Co., Ltd, Lot No:20190820)

4.0 Experiment design and dose

4.1 Sample preparation

According to the table above, 5 samples were randomly selected for the experiment.

4.2 Test method

Weigh each mask prior testing. The full mask is aseptically removed from the packaging and placed in a sterile 500 ml bottle containing 300 ml of extraction liquid (1 g/l Peptone, 5g/l NaCl and 2 g/l Tween 20). The bottle is laid down on an orbital shaker and shaken for 5 min at 250 rpm. After this extraction step, 100 ml of the extraction liquid is filtered through a 0,45 µm filter and laid down on a TSA plate for the total viable aerobic microbial count. Another 100 ml aliquot of the same extraction liquid is filtered in the same way and the filter plated on Sabouraud Dextrose agar (SDA) with chloramphenicol for fungi enumeration. The plates are incubated for 3 days at 30°C and 7 days at 25°C for TSA and SDA plates respectively. The total bioburden is expressed by addition of the TSA and SDA counts.



5.0 Statistical method

Count according to the principle of colony count.

6.0 Results of the test

Culture start time		2020-08-20 10:20			
Culture end time		2020-08-23 10:20	2020-08-27 10:20	Total bioburden (cfu/per sample)	Total biomass (cfu/g)
sample number	weight	Aerobe (cfu/100ml)	Fungi (cfu/100ml)		
1	3.4	1	< 1	< 6	< 2
2	3.3	1	< 1	< 6	< 2
3	3.3	4	2	18	5.5
4	3.3	< 1	< 1	< 6	< 2
5	3.4	< 1	< 1	< 6	< 2

According to EN 14683-2019 standard, the total biomass of surgical masks should be ≤30cfu/g.

7.0 Conclusion

Under the conditions of this study, the test article met the criteria.

8.0 Record

All raw data pertaining to this study and a copy of the final report are to be stored in the designated archive files at Huatongwei.

9.0 Confidentiality Agreement

Statements of confidentiality were as agreed upon prior to study initiation

Note:

This report is only responsible for the test result of submitted samples

* The inspection basis of mark * is not within the scope of CNAS recognition of the laboratory or the test results are provided to qualified outsourcing institutions.

Based on the report No.: 4478420080329modify, and replacing the original report No.: 4478420080329.

Approved by



Date:2020-09-08